NETWORK RECONSTRUCTION VIA DYNAMIC SYSTEMS: BIOLOGICAL NETWORK RECONSTRUCTION USING DIFFERENTIAL EQUATION MODELS James Henderson and George Michailidis jbhender@umich.edu, gmichail@umich.edu

PROBLEM: NETWORK RECONSTRUCTION

Gene regulatory networks represent functional relationships among genes as a directed network with edges corresponding to regulatory relationships; *i.e.* an edge $i \rightarrow j$ denotes that gene *i* regulates gene *j*. **Recovering the regulatory relationships among genes is a fun**damental problem in systems biology with important applications in disease treatment and diagnosis.

COMMON APPROACHES

Conditional Independence Models, including Bayesian Networks, are based on learning a *directed acyclic graph* encoding conditional independence relations. However, this approach cannot accomodate biologically relevant cycles such as feedback loops. Time Series Models including Dynamic Bayesian Networks and Vector Autoregressive Models avoid this limitation by restricting edges to point forward in time. **ODE** Models are perhaps the most widespread class of models in the biological and physical sciences. *ODE-based methods* for network reconstruction from time-series data have been proposed but take an ad-hoc approach to inference.

NETWORKS, DYNAMIC SYSTEMS, & COUPLING

This poster describes methodology for time series data, assumed to be noisy observations of an underlying dynamic system,

$$\dot{x}^{(r)}(t) = \boldsymbol{\alpha} + f(\boldsymbol{x}^{(r)}(t)) + \boldsymbol{u}^{(r)}(t), \quad \boldsymbol{y}^{(r)}(t_k) = \boldsymbol{x}^{(r)}(t_k)$$

with r indexing inedpendent experiments and $u^{(r)}(t)$ representing a vector of control parameters. Only the trajectories are observed, not the derivatives.

Network reconstruction is recast as variable selection for a **dynamic system**; i.e. determining the important variables in f,

$$i \to j \iff \frac{\partial f_i}{\partial x_j}(\boldsymbol{x}(t)) \not\equiv 0.$$

Rank potential edges using the **coupling metric**,

$$\rho(i,j) := \int_0^1 \left| \frac{\partial f_i}{\partial x_j}(\boldsymbol{x}) \right| d\boldsymbol{x},$$

measuring the strength of the regulatory relationship $i \to j$. Signed coupling metrics $\rho_+(i,j), \rho_-(i,j)$ can be defined similarly by replacing \cdot | with (\cdot)₊ or (\cdot)₋ respectively; useful for recovering signed edges.

REFERENCES

- [1] Gugushvili, Shota and Klaassen, Chris A.J. ' \sqrt{n} -consistent parameter estimation for systems of ordinary differential equations: bypassing numerical integration via smoothing.' In *Bernoulli*, 2012.
- Ravikumar, P., Lafferty, J., Liu, H. and Wasserman, L. 'Sparse Additive Models.' In *JRSS:B*, 2009.

 $_k) + \boldsymbol{\epsilon}_k, \quad (1)$

(3)

ODE ESTIMATION: A TWO-STAGE APPROACH

Stage 1: Estimate trajectories and derivatives using smoothing splines

- Avoids computationally intensive numerical integration
- Decouples the ODE estimation into d sub-problems
- Stage 2: Fit an additive, non-parametric, approximation to the ODE • The non-parametric approach allows for flexible nonlinear models and sidesteps potential model misspecification.
- - An additive approximation keeps the problem computationally feasible.

PERFORMANCE: E. Coli 2 SUBNETWORK



Edge and sign recovery on the E. coli 2 subnetwork. Line type is used to indicate edge recovery among the top 15-ranked edges: solid lines are true positives (10), dashed lines are false negatives (5), and dotted lines are false positives (5). Blunt-arrows indicate negatives on the data-generating network, while negatives on the estimated network are colored red. The area under the ROC curve and Precision-Recall curve are .87 and .68, respectively. All 15 signs are recovered correctly.

TUNING PARAMETERS

There are 3 sets of tuning parameters to be selected:

- λ_0 controls the smoothness of the first-stage estimates,
- λ_1 controls the smoothness of the additive components,
- λ_2 controls the sparsity of each additive model.

Leave-one-experiment-out cross validation is used to select $\lambda_2 = \lambda_{2i}$ while $\lambda_0 = \lambda_{0,ir}$ is selected by GCV. Here $\lambda_1 = .76$ is fixed.



STAGE 1: SMOOTH



$$\hat{x}_{i}^{(r)}(t) = \arg\min_{x \in W_{2}^{2}} n^{-1} \sum_{k=1}^{n} [$$

and $\hat{\dot{x}}_i(t) = \frac{d}{dt}\hat{x}_i(t)$.

Gene 2



derivative estimate from the first stage. \hat{f} Estimate the r

right-hand side function
$$f$$
 using $f_i = \arg\min M_{in}^{(r)}(f)$,
 $M_{in}(f) = ||\hat{x}_i - \sum_{j=1}^p f_{ij}(\hat{x}_i)||_n^2 + J(\lambda_1, \lambda_{2i}),$ (5)

$$J(\lambda_1, \lambda_2) := \sum_{j=1}^p \lambda_1 \int [\ddot{f}_{ij}(x)]^2 dx + \lambda_2 \sum_{j=1}^p \sqrt{\int [f_{ij}(x)]^2 dx}.$$
 (6)

algorithm (Ravikumar, 2009).



The estimated differential equation (dashed line) for gene 2 (left) and gene 5 (right) following a knockout of gene 1. The estimates are sums of the additive components (solid lines). The dotted lines show the $\cdot \pi(r)$

For knockout time-series constrain $f_{ii}(x_i) = \alpha 1_{[r=i]} + \beta x_i$. The minimization is carried out using a modified version of the sparse-backfitting