

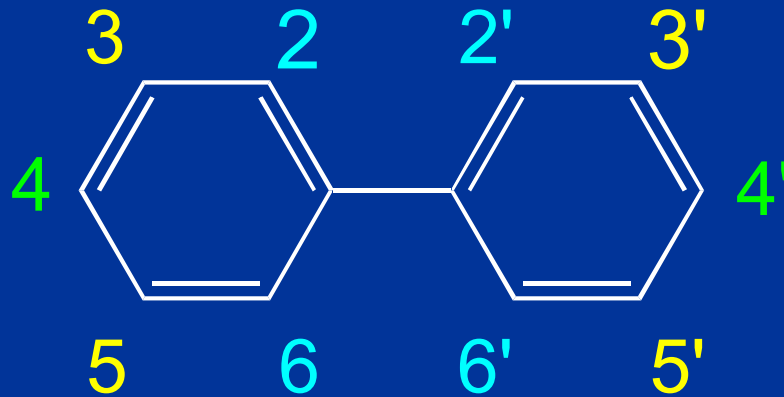
Stimulating Microbial  
Dechlorination of PCBs *in situ*

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# Polychlorinated Biphenyls (PCBs)



*ortho* 2, 6

*meta* 3, 5

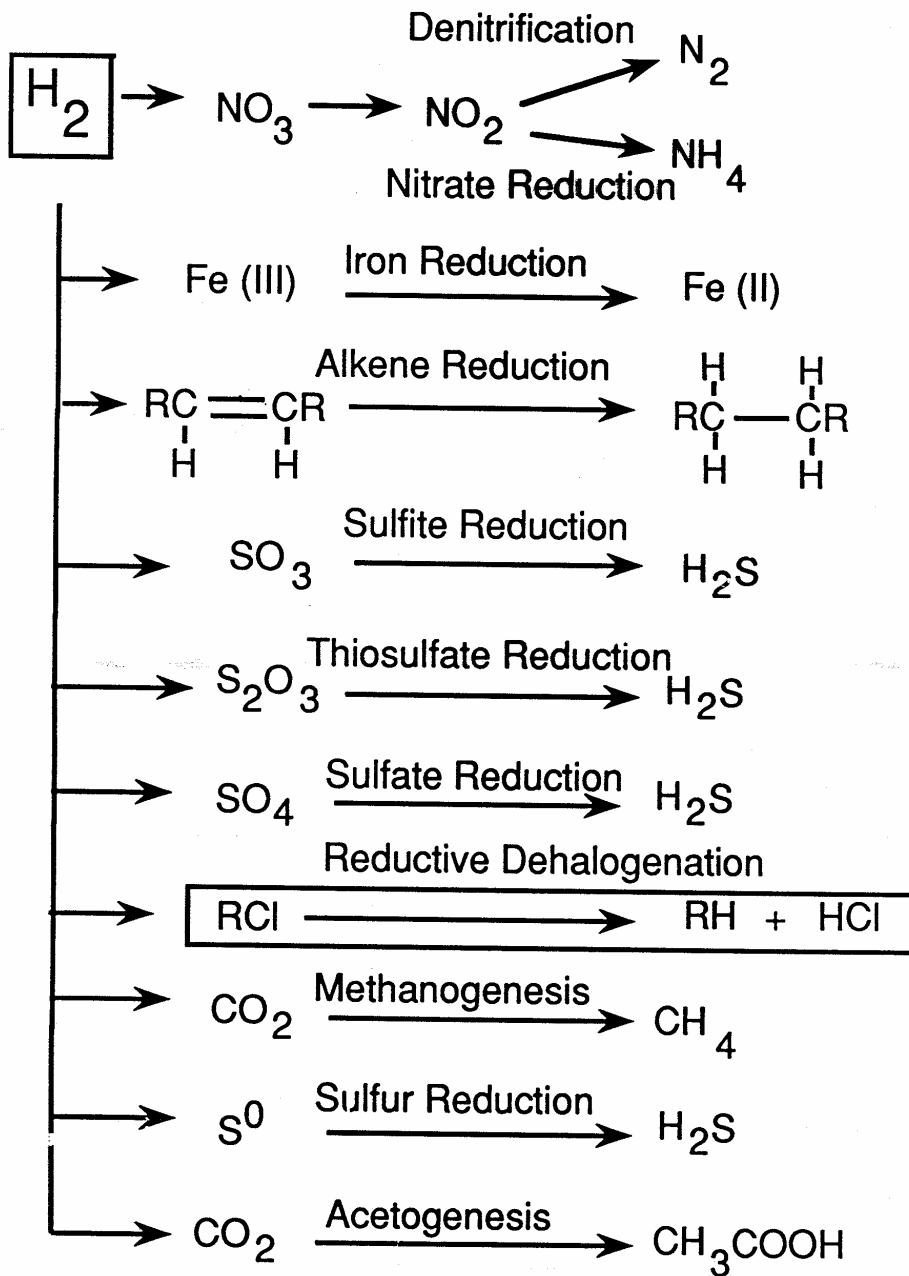
*para* 4

# U.S. Sites Containing PCB-Dechlorinating Microorganisms

LOCATION	PCB-CONTAMINATED SITE
Florida	Escambia Bay
Georgia	LCP Superfund Site (Brunswick)
Illinois	Waukegan Harbor
Maryland	Baltimore Harbor
Massachusetts	Hoosic River, Silver Lake, New Bedford Harbor Housatonic River / Woods Pond
Michigan	River Raisin
South Carolina	Charleston Harbor, Lake Hartwell
Washington	Puget Sound
Wisconsin	Fox River, Sheboygan Harbor and River

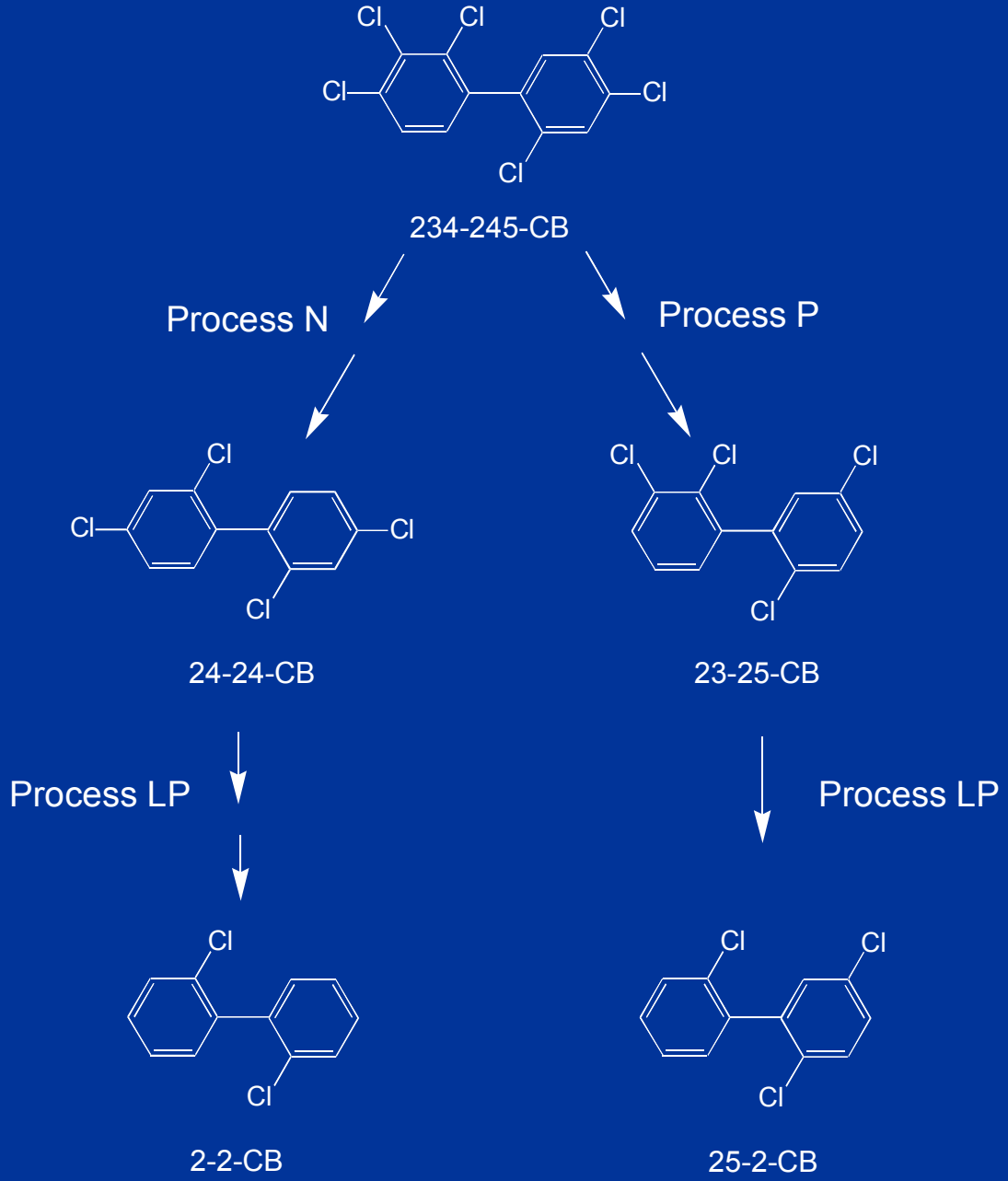
# What Influences Microbial PCB Dechlorination?

- PCB concentration
- PCB congener distribution
  - Penta- and Hexa-CBs, multiple flanked chlorines
  - Tri- and Tetra-CBs, few flanked chlorines
  - Hepta- Nona-CBs , less soluble, steric hindrance
- Environmental Conditions: Temperature, pH
- Electron donors: H<sub>2</sub>, organic compounds
- Electron acceptors: sulfate, iron, nitrate, carbonate
- Co-contaminants:
  - Oil, organics
  - Pesticides, xenobiotics
  - Heavy metals

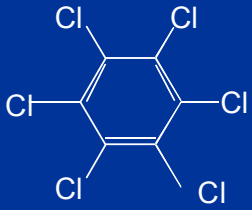


# Electron Accepting Reactions in Anaerobic Environments

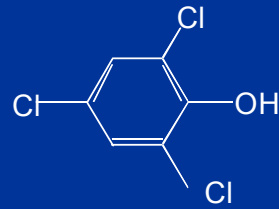
# Three Different Dechlorination Processes in Woods Pond: N, P, and LP



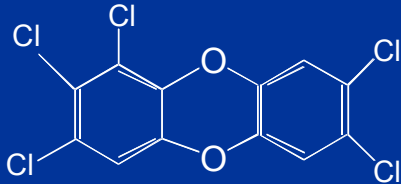
# Halorespiration



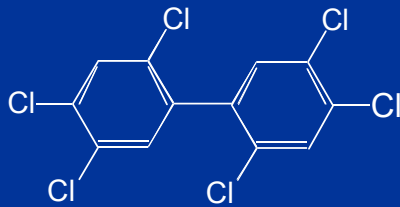
CHLOROBENZENE



CHLOROPHENOL



1,2,3,7,8-PeCDD



PCBs ?

- Some bacteria require halogenated aromatics for growth

# Priming PCB Dechlorination

## *Hypothesis*

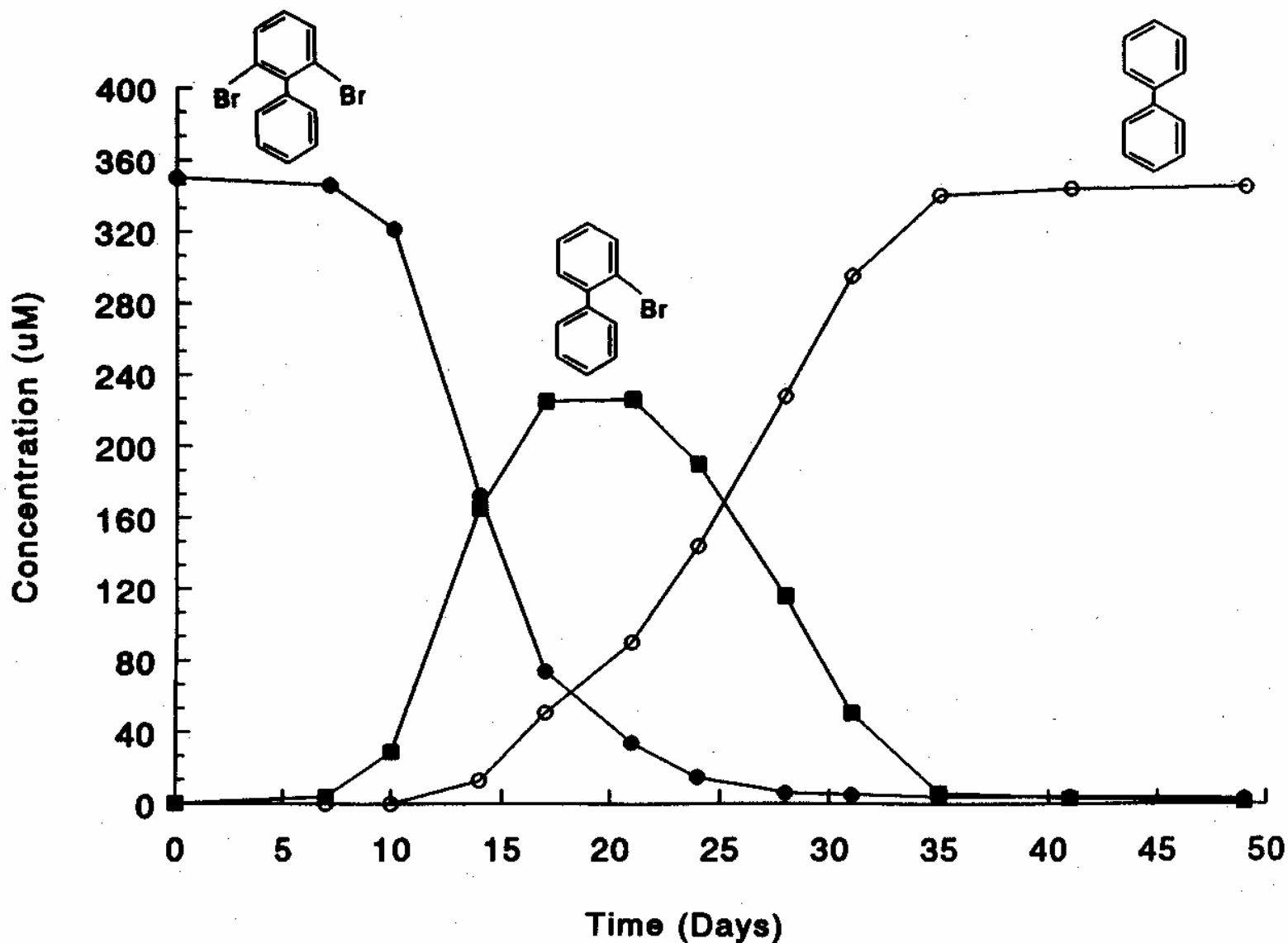
- PCB dechlorinators use halogenated biphenyls as electron acceptors

## *Corollary*

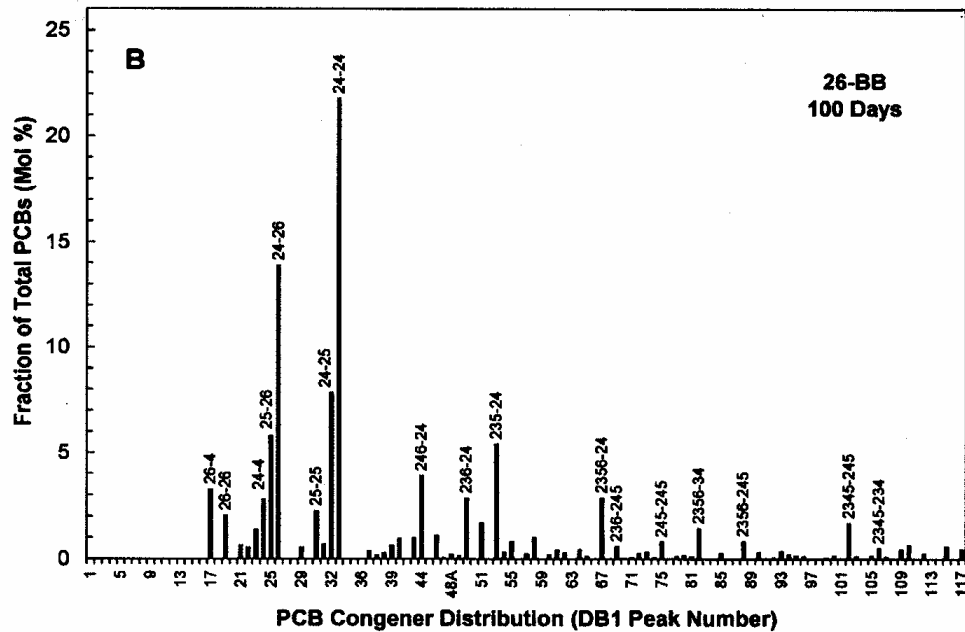
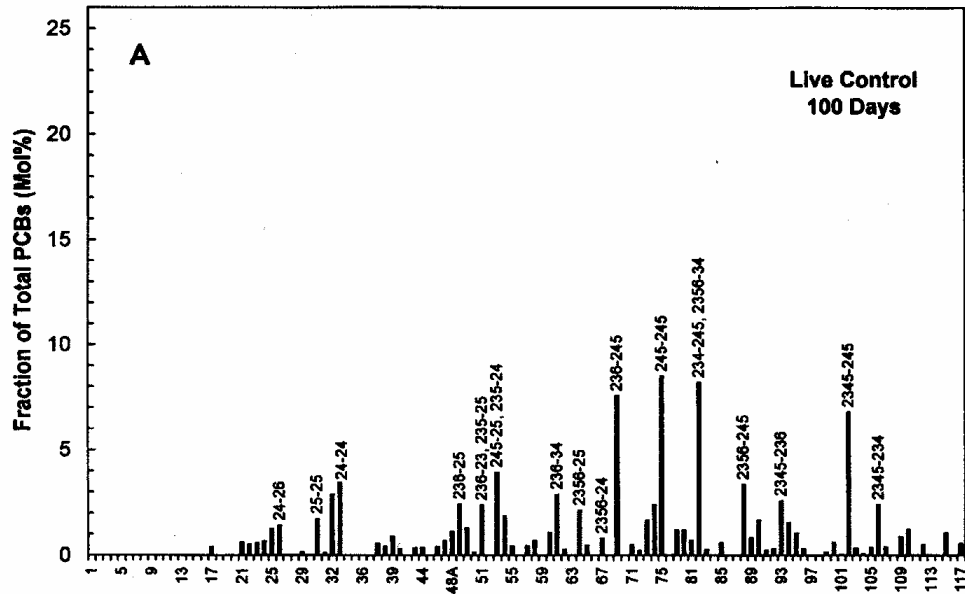
- Halogenated biphenyls can be used to selectively enrich PCB-dechlorinating bacteria



# Time Course of Dehalogenation of 26-BB



# Effect of Process N Dechlorination on PCB Congener Distribution



# Priming PCB Dechlorination

## *Hypothesis*

- PCB dechlorinators use halogenated biphenyls as electron acceptors

## *Corollary*

- Halogenated biphenyls can be used to selectively enrich PCB-dechlorinating bacteria

## *Result*

- 26-BB → 1000 fold increase in PCB dechlorinators (from  $10^5$  to  $10^8$  cells/ml)
- Increased numbers of PCB dechlorinators dechlorinate decades old PCBs in sediment

# Field Test Parameters

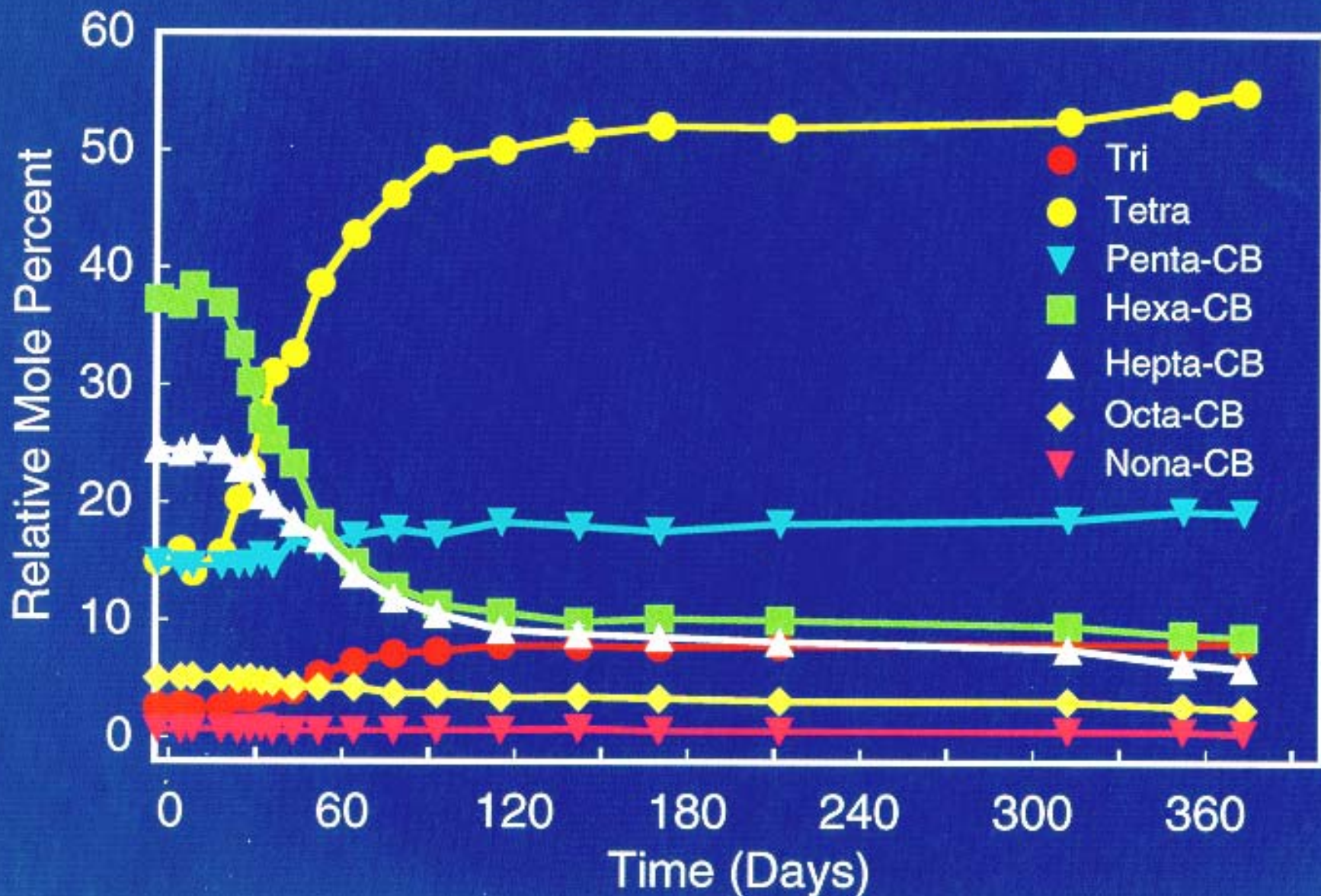
- 400 Kg Sediment (Dry weight)
- 3500 L Water and Sediment
- PCBs 26 ppm (dry wt)

## Amendments

- 2,6-dibromobiphenyl 350  $\mu\text{M}$  (880 ppm)
- 17 L Acetone (0.5%)
- Disodium malate (10 mM)

# Woods Pond Field Test

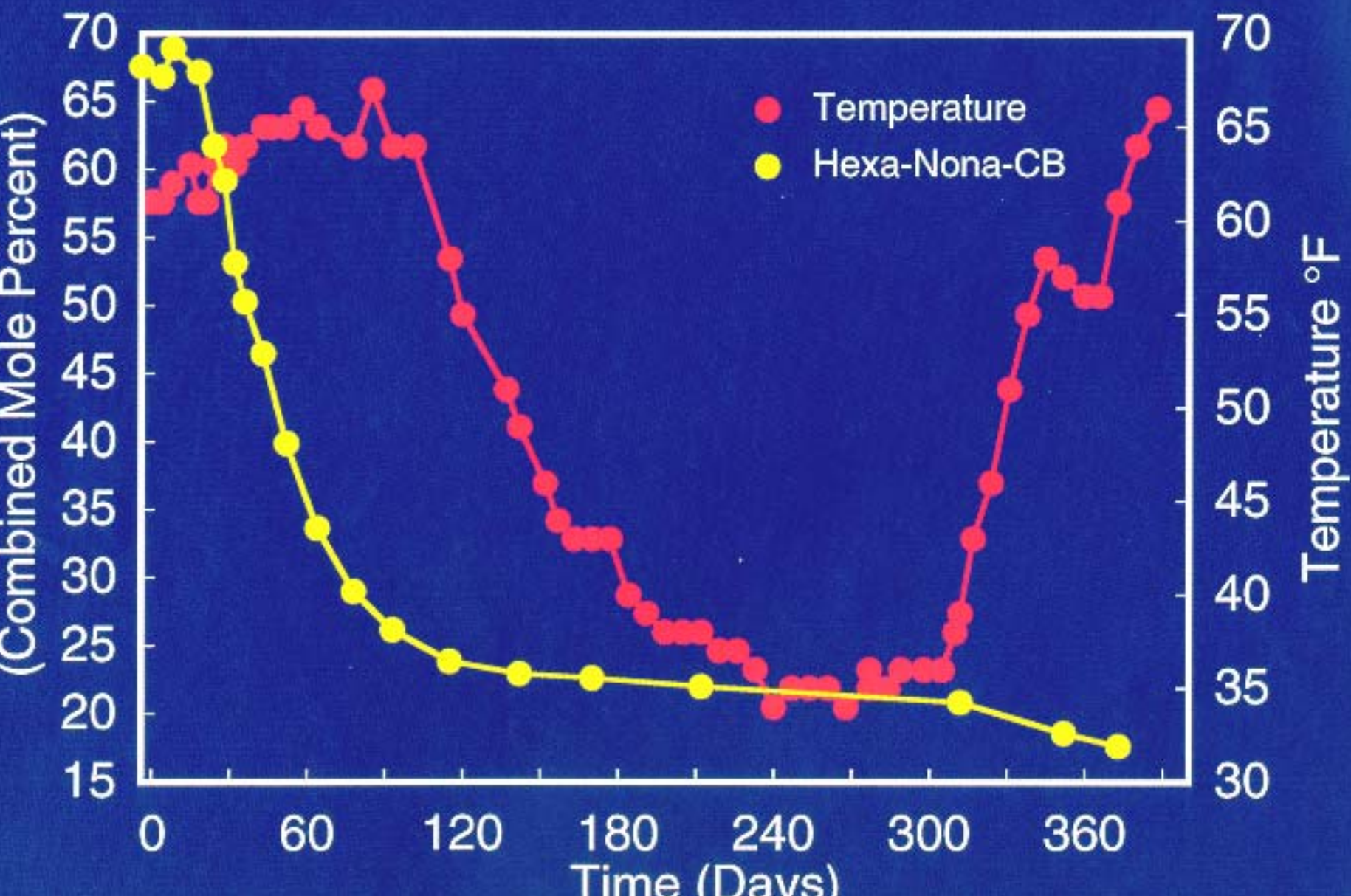
## Dechlorination of PCBs in Sediment, Top 6"





# Woods Pond Field Test

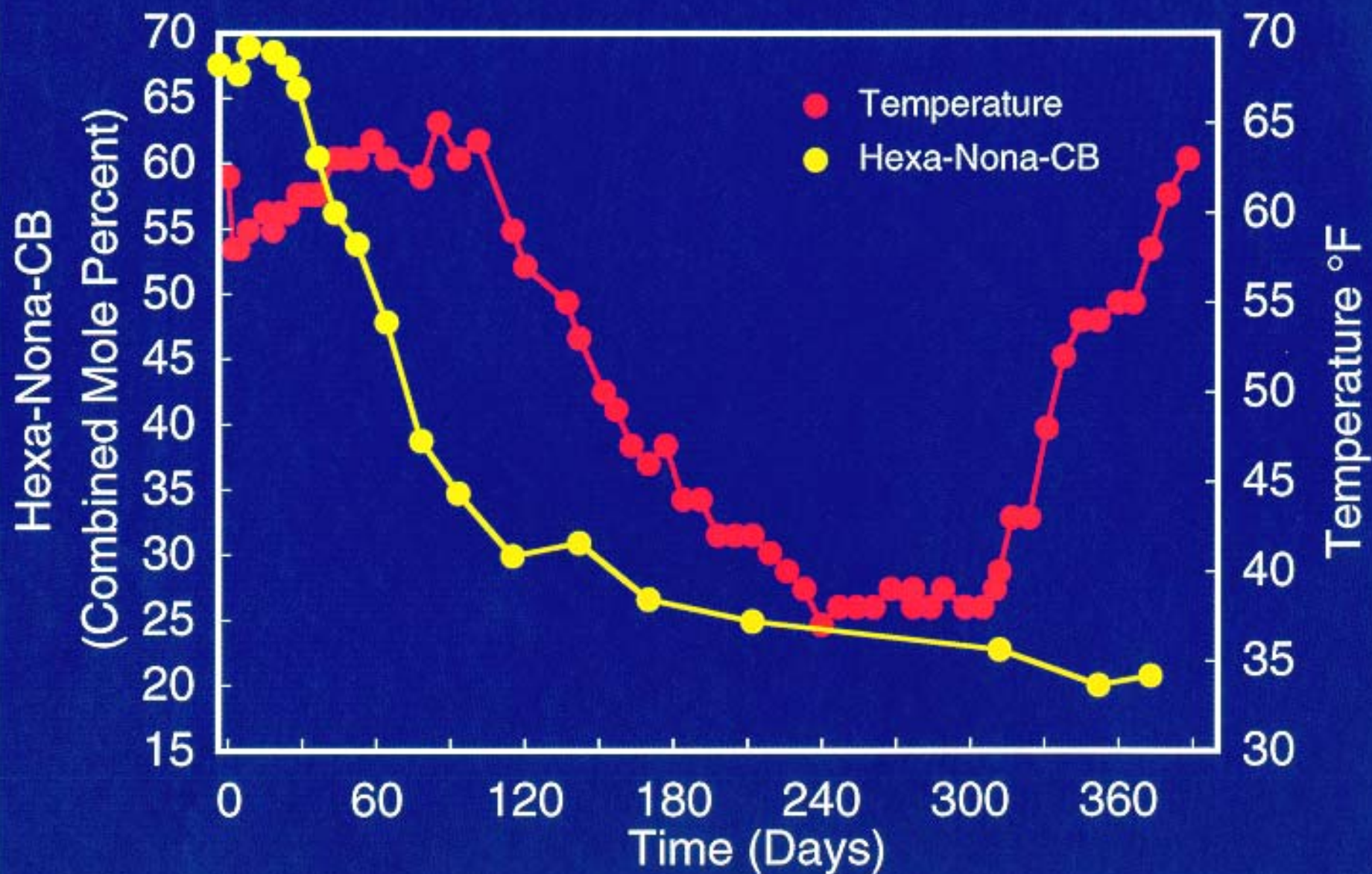
## Decrease in Hexa- to Nona-CB, Top 6" of Sediment



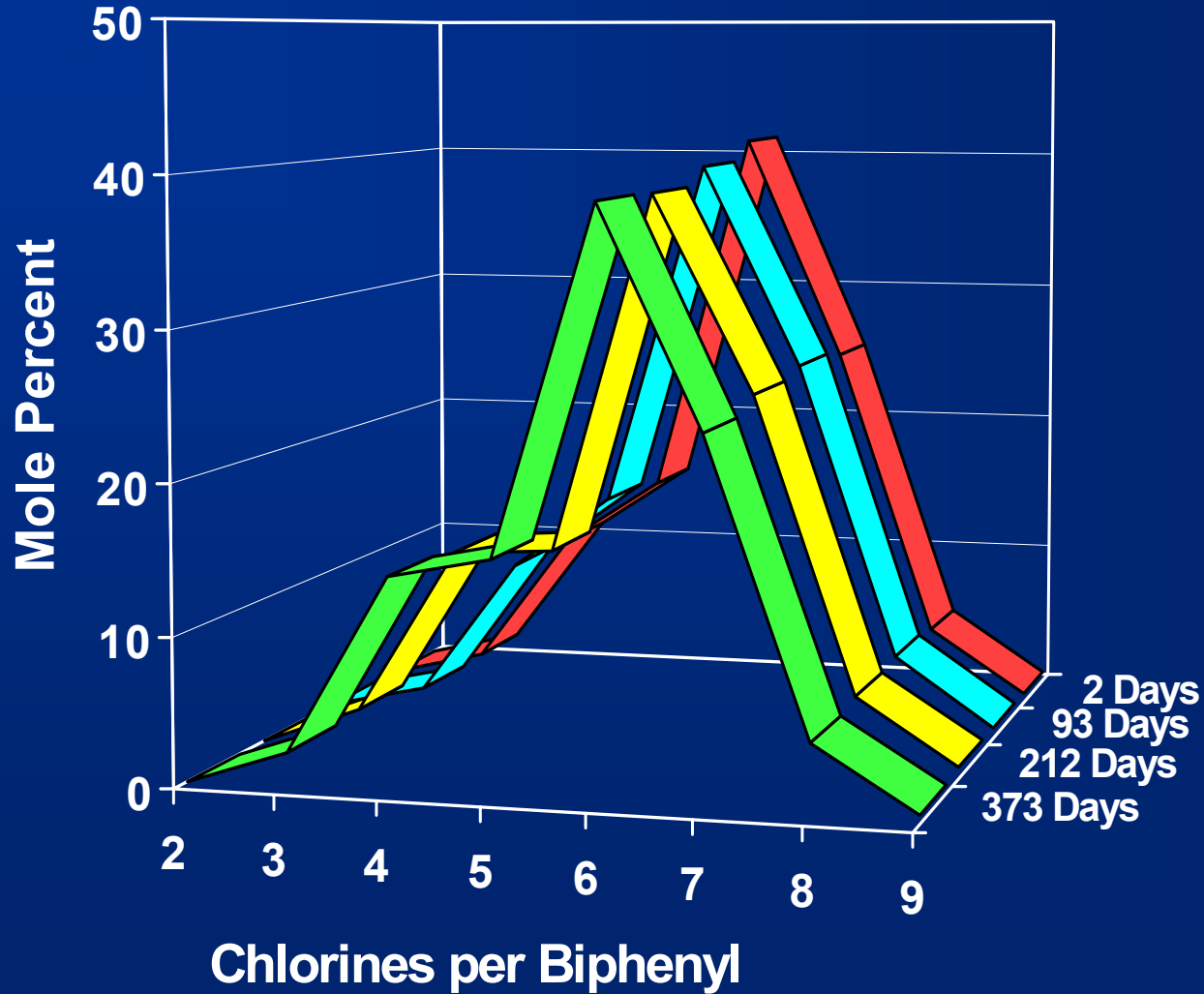


# Woods Pond Field Test

## Decrease in Hexa- to Nona-CB, Bottom 6" of Sediment

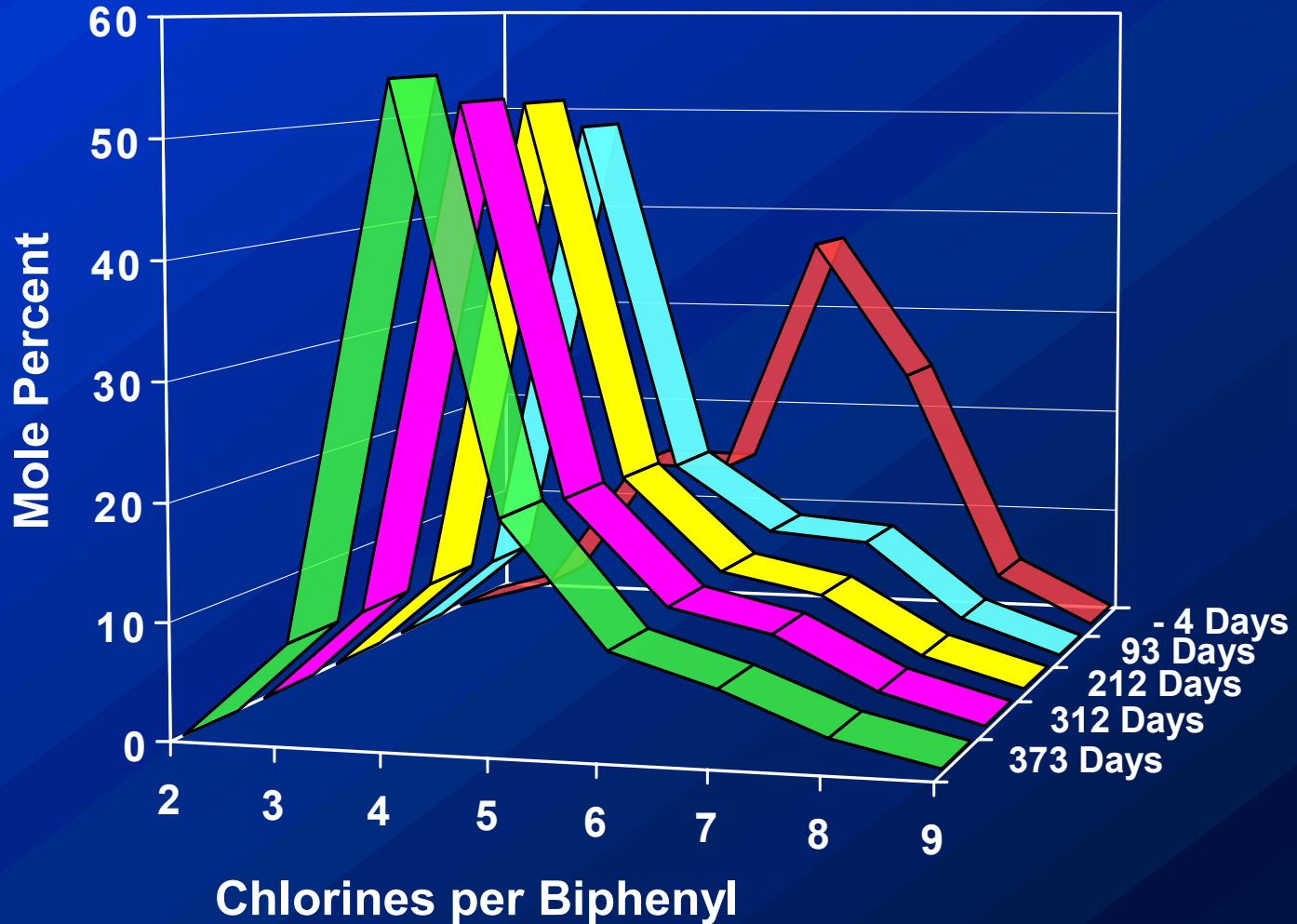


# PCB Homolog Distribution: Control Caisson





# PCB Homolog Distribution: Experimental Caisson



# Field Test Results

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Day	Hexa- to Nona-CBs	
	Mol %	% Decrease
0	68	
93	26	62
212	22	68
373	17.6	74

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# Field Test Results

Homolog	Initial Mol %	% Decrease	
		93 Days	373 Days
Hexa-CB	38	70	77
Hepta-CB	24	57	75
Octa-CB		27	50

# Strategy for Enriching/Isolating PCB Dechlorinating Bacteria

- Exploit specific temperature, primer, and pH requirements
- Use inhibitors and antibiotics to inhibit bacteria not essential to the PCB-dechlorinating community
- Repeatedly transfer with primers onto sterile PCB-contaminated sediment and then defined medium
- Use molecular tools to monitor the microbial population and identify key microorganisms

# Recent Findings

- Putative PCB dechlorinators in Woods Pond are *Dehalococcoides* spp.
- Grow on defined medium while retaining full PCB-dechlorinating activity
- *Dehalococcoides* genome reveals 17 reductive dehalogenase homologs (TIGR)
- Dehalorespiration is only known means of growth
- *Dehalococcoides* substrates:
  - Dhc sp.* CBDB1: Hexachlorobenzene, 1,2,3,7,8-PeCDD (Lorenz Adrian and colleagues)
  - Dhc ethenogenes* : Hexachlorobenzene, 1,2,3,4-TeCDD (Steve Zinder and Donna Fennel)

# Evaluation of the Technology

## Strengths

- Natural process
- Effective *in situ* at 12 to 30°C
- May be able to grow dechlorinators in lab and bioaugment without adding a primer
- Can reduce toxicity & persistence

## Weaknesses

- Success depends on microbial composition
- Affected by organic content and xenobiotics
- Requires addition of a halogenated primer
- May not work well for very low concentrations
- Dechlorinates but does not destroy

# Acknowledgments

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